



Documentation

CmiRClustFinder v2.0 Tool

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If you are using this pipeline, please cite:

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Example Dataset

To download example datasets (.zip), please click here

File Details:

File	Description
miRNA-Clusters.bed	Four-column simple bed file contain
	clustered miRNA and their genomic
	location
Segmented-SCNA.seg	Segmented SCNA from the TCGA-
	CHOL (cholangiocarcinoma) patients





Thoughts behind the development of the CmiRClustFinder Tool

Copy number aberration (CNA) events are prevalent in cancer patients. Recurrent and aberrant copy number regions can occur in genetic regulators such as microRNA (miRNA) clusters. The miRNA clusters are groups of miRNAs that are co-expressed, with similar expression patterns and jointly regulate target genes that are associated with various normal biological phenomena and tumor-associated pathways. Several studies have demonstrated CNA-mediated miRNA dysregulation; however, the interplay between clustered miRNAs and CNAs in cancer progression is largely unknown.

We hypothesize that the abnormal expression of clustered miRNAs due to aberrant CNAs and genomic rearrangements leads to the initiation and progression of several cancer types. Therefore, an understanding of miRNA clusters associated with CNAs could shed light on CNA-regulated tumor initiation and progression. Hence, we have developed a user-friendly computational pipeline, called CmiRClustFinder v1.0 by integrating multiple R-based and Linux command-line packages. CmiRClustFinder integrates CNA, gene and miRNA expression datasets from TCGA to compute CNA co-localized miRNA clusters, miRNAs, genes and user-defined genetic elements from 35 cancer types. The automated pipeline CmiRClustFinder v1.0 downloaded can be from the GitHub repository (https://github.com/msls-bioinfo/CmiRClustFinder_v1.0). The pipeline can be effectively used for integrated high throughput data analytics and identification of signatures for cancer diagnosis.

"CmiRClustFinder is not limited to only miRNAs/miRNA clusters, Users can use it for other genomic elements/regions."



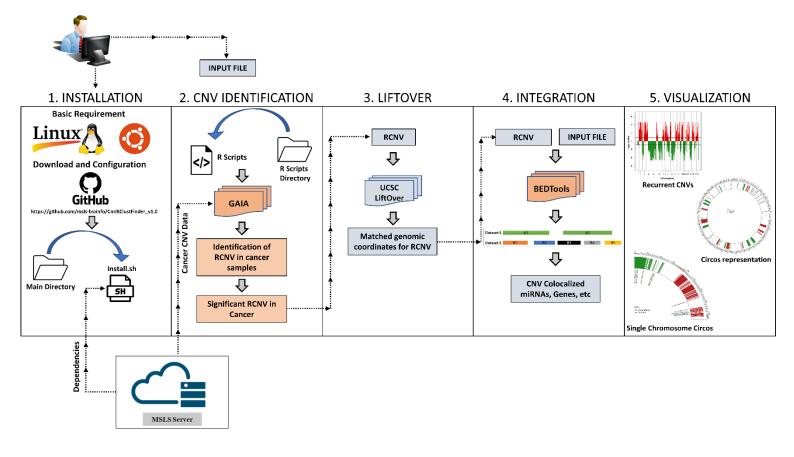


Standalone CmiRClustFinder v1.0 (A Linux based pipeline)





How does it work?



CmiRClustFinder v1.0 computes the integrated data within five steps. The installation script will download all the necessary resources and prepare the pipeline for use in the first step. In the second step, the GAIA package finds frequent aberrations in chromosomal regions among cancer patients' datasets. In the third step, the LiftOver tool matches the genomic build for RCNVs and user-defined genetic elements. We have integrated BEDTools to find co-localization of significant RCNV and genomic elements in the fourth step. Lastly, the circalize package generates a circos representation of the data.





Installation and Prerequisite

CmiRClustFinder pipeline is designed for Linux operating system. If you wish to use the standalone version of this pipeline, follow the instructions below.

The following Linux utilities are required to run this pipeline. Please make sure the following are installed and available on your system prior to running install.sh from the source directory.

- 1. R = 4.0 (or higher)
- 2. git
- 3. unzip

If the above prerequisites are satisfied, you are ready to install dependencies and build the program. Note during the building procedure, install.sh will attempt to download and install several packages, so an active internet connection is required.

To obtain CmiRClustFinder, Use:

git clone <u>https://github.com/msls-bioinfo/CmiRClustFinder_v1.0.git</u> cd CmiRClustFinder_v1.0/ Or wget <u>https://github.com/mslsbioinfo/CmiRClustFinder_v1.0/archive/refs/heads/main.zip</u> unzip main.zip cd CmiRClustFinder_v1.0-main/

If you have downloaded the source code and it is in a directory CmiRClustFinder/, to install all dependencies follow the procedure

cd CmiRClustFinder/ sh install.sh





Running the pipeline

After the successful execution of install.sh you are ready to run the main pipeline script CmiRClustFinder.r which is in RScript/ directory

Please navigate in to the Rscripts/ directory to start the pipeline

General usage:

Rscript CmiRclustFinder.r <TCGA cohort abbreviation> <BED file specifying user interested genomic regions>

Arguments / Parameters	Description					
<tcga abbreviation="" cohort=""></tcga>	This argument required TCGA cohort					
	abbreviation, which you can select from the list					
	below					
<i><bed file="" genomic<="" i="" interested="" specifying="" user=""></bed></i>	BED file which contains the specific genomic					
regions>	regions, to check their co-localization with					
	RCNV.					



TCGA Cancer Abbreviations

Sr. No.	Cohort Abbreviation	Cohort Name
1	TCGA-ACC	Adrenocortical carcinoma
2	TCGA-BLCA	Bladder Urothelial Carcinoma
3	TCGA-BRCA	Breast invasive carcinoma
4	TCGA-CESC	Cervical squamous cell carcinoma and endocervical adenocarcinoma
5	TCGA-CHOL	Cholangiocarcinoma
6	TCGA-COAD	Colon adenocarcinoma
7	TCGA-COADREAD	Colorectal adenocarcinoma
8	TCGA-DLBC	Lymphoid Neoplasm Diffuse Large B-cell Lymphoma
9	TCGA-ESCA	Esophageal carcinoma
10	TCGA-GBM	Glioblastoma multiforme
11	TCGA-GBMLGG	Glioma
12	TCGA-HNSC	Head and Neck squamous cell carcinoma
13	TCGA-KICH	Kidney Chromophobe
14	TCGA-KIPAN	Pan-kidney cohort (KICH+KIRC+KIRP)
15	TCGA-KIRC	Kidney renal clear cell carcinoma
16	TCGA-KIRP	Kidney renal papillary cell carcinoma
17	TCGA-LGG	Brain Lower Grade Glioma
18	TCGA-LIHC	Liver hepatocellular carcinoma
19	TCGA-LUAD	Lung adenocarcinoma
20	TCGA-LUSC	Lung squamous cell carcinoma
21	TCGA-MESO	Mesothelioma
22	TCGA-OV	Ovarian serous cystadenocarcinoma
23	TCGA-PAAD	Pancreatic adenocarcinoma
24	TCGA-PCPG	Pheochromocytoma and Paraganglioma
25	TCGA-PRAD	Prostate adenocarcinoma
26	TCGA-READ	Rectum adenocarcinoma
27	TCGA-SARC	Sarcoma
28	TCGA-SKCM	Skin Cutaneous Melanoma
29	TCGA-STAD	Stomach adenocarcinoma
30	TCGA-TGCT	Testicular Germ Cell Tumors
31	TCGA-THCA	Thyroid carcinoma
32	TCGA-THYM	Thymoma
33	TCGA-UCEC	Uterine Corpus Endometrial Carcinoma
34	TCGA-UCS	Uterine Carcinosarcoma
35	TCGA-UVM	Uveal Melanoma





The second argument required for Rscript is the BED file which contains the specific genomic regions, to check their co-localization with RCNV. Below is an example of the BED file.

NOTE: The table header is for descriptive purposes, the BED file should not have a header

CHROM	START	END	IDENTIFIER
chr19	53666679	53706336	hsa-miR-526a-1/miR-512-1
chr14	1.01E+08	1.01E+08	hsa-miR-1185-1/miR-379
chr14	1.01E+08	1.01E+08	hsa-miR-136/miR-493
chrx	50003148	50014683	hsa-miR-502/miR-532
chr9	1.35E+08	1.35E+08	hsa-miR-3689f/miR-3689c
chr13	91350605	91351391	hsa-miR-92a-1/miR-17
chrx	1.34E+08	1.34E+08	hsa-miR-106a/miR-363
chrx	1.35E+08	1.35E+08	hsa-miR-424/miR-450b
chrx	1.46E+08	1.46E+08	hsa-miR-891b/miR-892c
chr20	63919449	63919939	hsa-miR-941-5/miR-941-1

Know more about the bed file format: (<u>http://genome.ucsc.edu/FAQ/FAQformat#format1</u>)





CmiRClustFinder v2.0

(A Webserver with GUI)





Module for analyzing user input data

CmiRClustFinder 2.0 is the upgraded version with a graphical user interface (GUI) for non-

Linux users.

Data Submission

The data submission form on the CmiRClustFinder webserver

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 A pipeline developed to identify Recurre	CmiRClustFinder v2.0		efined datasets
Analyze Your Data TCGA Analysis Downloa	ad Pipeline	stFinder Modules	Documentation
erview is user-friendly WebTool is GUI for CmiRClustFinder pipelin upload a CNV File (.txt): Choose file No file chosen Upload Upload Genetic elements (.bed): Choose file No file chosen Upload	e, which identefies recurrent copy number variatie Segmented somatic copy number alterations (sCNA) data BED file which contains the specific genomic regions, to check their co- localization with RCNV.	ns and integrates user-inputted genor	mic element (eg. any specific region or

Dataset Example

Segmented copy number data

Sample Chromosome St	art End	Num Probes	Segment Mea	n	
TCGA-3X-AAV9-10A-01D-A419-		3218610	247813706	12909	6 0
TCGA-3X-AAV9-10A-01D-A419-		484222	9948475	5699	0.0088
TCGA-3X-AAV9-10A-01D-A419-		9949576	9950371	3	-1.5053
TCGA-3X-AAV9-10A-01D-A419-		9953949	70256255	35199	
TCGA-3X-AAV9-10A-01D-A419-		70258987	70259495	3	-0.9245
TCGA-3X-AAV9-10A-01D-A419-		70263331	76769258	3626	0.0022
TCGA-3X-AAV9-10A-01D-A419-		76789156	76799126	6	-1.014
TCGA-3X-AAV9-10A-01D-A419-	01 2	76808048	222751806	74929	0.005
TCGA-3X-AAV9-10A-01D-A419-	01 2	222754101	222759558	4	-1.11
TCGA-3X-AAV9-10A-01D-A419-	01 2	222760279	242476062	11782	0.0028
TCGA-3X-AAV9-10A-01D-A419-	01 3	2212571	10559238	4761	0.0039
TCGA-3X-AAV9-10A-01D-A419-	01 3	10560942	10562533	2	-1.5264
TCGA-3X-AAV9-10A-01D-A419-	01 3	10565125	65047603	31780	0.002
TCGA-3X-AAV9-10A-01D-A419-	01 3	65047633	65048037	2	-1.3193
TCGA-3X-AAV9-10A-01D-A419-	01 3	65051762	65432030	333	-0.0053
TCGA-3X-AAV9-10A-01D-A419-	01 3	65432464	65432483	2	-1.1371
TCGA-3X-AAV9-10A-01D-A419-	01 3	65434647	80933596	9483	-0.0029
TCGA-3X-AAV9-10A-01D-A419-	01 3	80935171	80936580		-1.267
TCGA-3X-AAV9-10A-01D-A419-	01 3	80940139	197538677	59974	0.0029
TCGA-3X-AAV9-10A-01D-A419-	01 4	1053934	36536256	21807	0.001
TCGA-3X-AAV9-10A-01D-A419-	01 4	36540003	36540033	2	-1.44

Bed file

chr16	16300159	16309966	hsa-miR-3180-2/miR-3179-2
chr16	18402178	18411977	hsa-miR-3179-3/miR-3180-3
chr1	220117853	220118241	hsa-miR-194-1/miR-215
chr12	69584722	69584822	hsa-miR-3913-2/miR-3913-1
chr16	14303967	14309371	hsa-miR-365a/miR-193b
chr16	14925937	14930879	hsa-miR-6770-1/miR-6511a-1
chr16	16324588	16329364	hsa-miR-6770-2/miR-6511a-2
chr16	18488301	18494576	hsa-miR-3179-4/miR-3670-4
chr18	21825698	21829088	hsa-miR-1-2/miR-133a-1
chr20	34990376	34998472	hsa-miR-4990/miR-499a
chr3	160404588	168484825	hsa-miR-16-2/miR-15b
chr9	124692442	124693798	hsa-miR-181b-2/miR-181a-2
chr9	128244721	128245030	hsa-miR-3154/miR-199b
chr9	128392618	128392708	hsa-miR-219b/miR-219a-2
chr22	46112749	46113768	hsa-let-7b/let-7a-3
chr8	12719132	12727299	hsa-miR-3926-2/miR-5692a-2
chr1	51059837	51060103	hsa-m1R-6500/m1R-4421
chr1	62078786	62078856	hsa-miR-3116-2/miR-3116-1
CULLA .	01010100	02070030	1158-111X-3110-2/11X-3110-





Module for analyzing user input genetic element against TCGA patient datasets

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A	pipeline developed to ide		ClustFinder v2.0 er Variations (RCNV) - colocalized		CGA & user defined	d datasets	
Analyze Your Data	TCGA Analysis	Download Pipeline				Documentation	
Overview Select cancer ty Select a cancer Upload BED File Choose file No Submit & Start A	file chosen	v	4	Upload	BED file v	GA cancer type fr which contains th with appropriat	e specific





Data Processing

After submitting the data in the proper format, it will be redirected to the data processing window. The time required for data processing depends on the size of data files submitted and the processing load on the server.

Data processing page, which will be redirected to results after completion







Interpreting CmiRClustFinder Output

The output from CmiRClustFinder contains Circos plots for the representation of genomic regions/RCNV regions/their colocalization. The. tsv master file will provide specific details of each genetic element that overlap with significant RCNVs from the specified dataset.

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	A pipeline develop	CmiRClustFinder V. ad to identify Recurrent Copy Number Variations (RCNV) - coloc			GA & user define	d datasets		
Results	Documentation							
	result of your analysis inder pipeline, which identefi	es recurent copy number variations and integrates user-inputted Click on icons to visualize/dowload results	l genomic elemer	nt (eg. any spe	cific region or ge	ne).		
	File Description Visualize/Download							
	Circos genetic elements PNG)	Circos represented of user provided genetic elements		POF				
C	Circos (PDF)	Circos represented of recurrent copy number variation or genetic elements	o-localized	x				
R	Result (TSV)	Circos represented of recurrent copy number variation or genetic elements	o-localized	ار				
R	Result (ZIP)	Compressed file of results		ZIP				
Go	Back							





Credits

- 1. TCGA Workflow: Analyze cancer genomics and epigenomics data using Bioconductor packages (<u>https://f1000research.com/articles/5-1542</u>)
- 2. TCGAbiolinks R package (<u>https://bioconductor.org/packages/release/bioc/html/TCGAbiolinks.html</u>)
- 3. BEDTools (<u>https://bedtools.readthedocs.io/en/latest/</u>)
- 4. UCSC liftOver (<u>https://genome-store.ucsc.edu/</u>)